

Comparison of Methods for Diagnosing Bacterial Vaginosis

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ABSTRACT

Objective: To determine the laboratory method that best predicts Bacterial Vaginosis.

Study Design: Descriptive Observational study

Place and Duration of Study: This study was conducted in the Department of Microbiology, Sindh Medical College (DUHS) and Basic Medical Sciences Institute, Jinnah Postgraduate Medical Centre, Karachi from Jan. 2005 to Feb. 2007.

Materials and Methods: A total of 150 randomly selected women were included in this study from OPD of various tertiary care hospitals and & family planning clinics of the city. In this study we compared and calculated the sensitivity, specificity and predictive value of positive and negative tests for wet mount, Gram stained vaginal smears and Gardnerella vaginalis (G.vaginalis) cultures with clinical sign Amsel's criteria (Gold standard), for the diagnosis of bacterial vaginosis(B.V).

Results: We diagnosed 54 (36%) cases of bacterial vaginosis by Gram's staining method, 61 (41.7%) cases by Wet Mount method, and 42 (28.0%) cases by Amsel's criteria and 47 (31.3%) cases by culturing.

Conclusions: Amsel's criteria were comparable with other laboratory tests for diagnosis of BV. Culture was laborious, expensive and least sensitive method.

Key Words: Bacterial vaginosis, Amsel's clinical criteria, Gram staining

INTRODUCTION

Bacterial vaginosis (BV) was initially recognized as a vaginal syndrome by Gardner and Duke¹. They associated bacterial vaginosis with the isolation of Haemophilus vaginalis, later named it Corynebacterium vaginale and currently named Gardnerella vaginalis². However, the microbiology of bacterial vaginosis is complex and involves organisms other than G.vaginalis. Large quantities of not only G.vaginalis but also anaerobic bacteria^{3,4,5} and Mycoplasma hominis^{3,4,6} can be recovered from women with bacterial vaginosis.

BV is the most common cause of malodorous vaginal discharge in females of child bearing age.^{6,9,11} BV is characterized by alterations in vaginal flora. Normally, Lactobacilli constitute 95% of bacteria in the vagina, but in BV, Lactobacilli are absent or severely reduced and the concentration of other bacteria is increased by 10^2 - 10^4 . The patho physiology of this syndrome is better understood, but little progress has occurred in identifying the casual factors.^{8,12} Now BV is increasingly recognized as directly related to a number of serious obstetric and gynecological complications.^{5,7,10}

G.vaginalis can be isolated from the vagina of 20% to 40% of women without bacterial vaginosis^{1,4,6}. Symptoms of bacterial vaginosis are nonspecific, and has been associated with severe sequelae^{13,14} and diagnosis should rely on confirmatory tests^{15,16,17}

MATERIALS AND METHODS

A total of 150 randomly selected women were included in this study. They were grouped as: 50 non-pregnant females complaining of discharge. 50 pregnant females, complaining of discharge. The third group included 50 Pregnant / Non-pregnant females not complaining of discharge (controls). The control group who had no sign and symptoms of vaginal discharge, was examined and studied in an identical manner. They were matched for age and socioeconomic status. All females with history of previous abortion, preterm delivery or premature rupture of membrane and infertility were included. Females attending family planning clinic were also included. The exclusion criteria were females taking antibiotic, using vaginal douches, tablets or suppositories within the preceding 14 days. Women who had sexual intercourse within 24 hours were also excluded^{18,19}

RESULTS

The results for the diagnosis of bacterial vaginosis by Amsel's criteria, culture, Wet Mount and Spiegel's criteria (Table 1). Statistical analysis showed that all the 4 methods could be used as a means for the diagnosis of bacterial vaginosis ($p < 0.01$).

Table I: Shows different methods employed for the diagnosis of BV. In both the groups the wet mount for clue cells was more diagnostic 61(41.7%). In group not complaining of vaginal discharge it was 19 (38%) and in

group complaining it was 42 (43%). The second method was Gram's method which was 37 (37%) in females complaining of discharge and 17 (34%) in females not complaining of discharge. So over all it was present in 54 (36%). The culture was least diagnostic and was only 31.3% diagnostic.

Table No.1: Methods used for Diagnosis of Bacterial Vaginosis (n=150)

Patients	Amsel's Criteria (Gold standard)	Wet Mount	Gram's Staining	Culture (HBT)
Not complaining vaginal discharged (n=50)	08 (16.0%)	19 (38.0%)	17 (34.0%)	16 (32.0%)
Complaining vaginal Discharge (n=100)	34 (34.0%)	42 (43.0%)	37 (37.0%)	31 (31.0%)
Total (n=150)	42 (28.0%)	61 (41.7%)	54 (36.0%)	47 (31.3%)

Table 2: Shows the comparison of the sensitivity, specificity, positive and negative predictive value of wet mount, gram staining and culture as compare to Amsel's criteria. According to this the positive predictive value, negative predictive value and the sensitivity and specificity of Wet Mount were 62.3%, 95.5%, 90.5% and 78.7% respectively. Those of Gram's staining were 72.2%, 96.9%, 92.9% and 86.1%

Table No.3: Reliability, Time Consumption and Approximate Cost of the Test Method for Detection of BV

Test Method	Reliability		Time Consumption	Cost Per Test	Labor
	Sensitivity (%)	Specificity (%)			
Amsel's	Standard or Reference Method		3 Min	Cheap	Very easy to perform
Wet Mount	90.5	78.7	5 Min	Cheap	Easy to perform
Gram's Staining	92.9	86.1	10-15 Min	Cheap	Easy but requires experience
Culture	64.3	81.5	24-72 Hrs	Costly	Laborious

A thin, homogenous, foul smelling discharge that is adherent to the vaginal walls is characteristic of B.V. The discharge should not be confused with cervical mucus, which is characteristically clear, indicating the absence of an inflammatory response. A milk like consistency that is distinctly nonflocular, nongranular, nonstringy and not clumped is most characteristic. The discharge is clear to grey in color but has occasionally been reported as green, yellow or even white^{1,4,18}. The volume of discharge varied from scanty, moderate to profuse. These criteria were used to define a normal or abnormal discharge (bacterial vaginosis) in all subsequent results and analysis.¹⁸

A detailed clinical history of each woman was taken and their two high vaginal swabs were collected. One swab was suspended in a sterile tube containing 0.5 ml

respectively. The culture was 64.3% sensitive and 81.5% specific, the positive predictive value was 57.4% and the negative predictive value was 85.4%.

Table No. 2: Sensitivity, Specificity, Positive and Negative Predictive value of wet mount, gram staining and Cultures as compare to amsel's criteria

	Wet mount	Gram's staining	Culture
Sensitivity	90.5%	92.9%	64.3%
Specificity	78.7%	86.1%	81.5%
Positive Predictive Value	62.3%	72.2%	57.4%
Negative Predictive Value	95.5%	96.9%	85.4%

Table 3: Shows if the four diagnostic methods were compared for the reliability, time consumption and approximate cost per test and labor to perform these tests. The Amsel's criteria was easy to perform and cheap method as compared to culture method which is time consuming, costly and quite labor intensive.

of sterile physiological saline and second swab was suspended in Stratus transport medium to be used for culture. The vaginal swabs were used for gram staining, for the determination of the pH of the vagina and for the Whiff test. Diagnosis of bacterial vaginosis was done by Amsel's criteria, Wet Mount Gram staining and by culture. The parameters that are necessary to decide the efficacy of the diagnostic tests, namely positive predictive value, negative predictive value and sensitivity and specificity were calculated in comparison with Amsel's criteria by considering it as the gold standard. Statistical analysis was done by using the Chi Square test. In all statistical analysis, only P values < 0.05 were considered to be significant.

Diagnosis by Amsel's criteria:

Amsel's composite criteria includes the presence of a homogeneous vaginal discharge, pH of the vagina being > 4.5, the presence of clue cells in gram stained vaginal discharge smears and a positive whiff test. According to Amsel, if 3 of the 4 criteria are positive, the patient has bacterial vaginosis^{4,20}

Vaginal pH determination:

Vaginal secretion or discharge was collected from the lateral vaginal walls with a cotton swab and this was then transferred onto a strip of pH paper. This was compared with a standardized colorimetric reference chart to estimate the actual pH²¹.

Whiff test:

A drop of vaginal discharge was mixed with a drop of 10% potassium hydroxide which was taken on a slide. A fishy smell indicated a positive test²²

Processing of sample:

The time period between collection of sample and inoculation was restricted to 1 hour.

Wet Mount Examination:

One drop of sterilized saline suspension was applied on a glass slide and covered with slip. It was examined microscopically for clue cells (vaginal epithelial cells with characteristic stippled or granulated appearance) that is the vaginal epithelial cells with indistinct cell border obscured by the large number of coccobacilli.^{19,23}

Lactobacilli were recorded as the predominant flora on wet mount if long morphologic types were judged to be the predominant form.

Clue cells:

The vaginal discharge was smeared on clean glass slides, air dried, heat fixed and stained by Gram's method. The vaginal epithelial cells which were completely covered by the gram variable coccobacilli so that their edges which normally have a sharply defined cell border became indistinct or stippled, were considered as the clue cells²⁴.

Diagnosis by culture:

The vaginal swabs were inoculated on selective differential Human Blood Bilayer agar medium with Tween 80 (HBT) culture media and incubated at 37°C for 24 to 48 hrs in a candle jar to provide 5-10% CO₂ (Totten et al 1982). * Aerobes, facultative anaerobes and obligate anaerobes were identified by their colony morphologies, gram staining and standard biochemical reactions.^{25,26} Those women of whom the culture showed predominant growth of *G. vaginalis* or an anaerobe or both were considered as positive for bacterial vaginosis by culture.

Diagnosis by Spiegel's criteria:

When the gram staining showed predominance (3 to 4+) of the lactobacillus morph type with or without the

Gardnerella morph type, it was interpreted as normal. When the gram staining showed a mixed flora consisting of gram-positive, gram negative, or gram-variable bacteria and the lactobacillus morphotype was decreased or absent (0 to 2+), the gram staining was interpreted as consistent with bacterial vaginosis.^{4, 21}

DISCUSSION

The goal of this study was to evaluate and correlate several clinical and microbiologic criteria that have been used for the diagnosis of BV. We were particularly interested in the diagnostic values of simple observations and procedure that could be carried out in the physician's office, and in the correlation between such office procedures and less costly and more readily available microbiologic test for BV.

In prior reports, individual laboratory methods of diagnosing bacterial vaginosis have been compared with clinical signs^{27,18,28,29}. However, multiple laboratory methods have not been compared with a single cohort of women. In the present study, we determined the vaginal flora of patients with bacterial vaginosis diagnosed by clinical signs, wet mount, gram stained vaginal smears and culture to document that each diagnostic method was associated with similar vaginal flora.

Results of this study agree with that of Amsel that majority of women who participated in the study remained free of any definite symptoms.^{18,7}

In this study, each of Amsel's clinical criteria (homogenous discharge, positive whiff test, vaginal pH>4.5 and clue cells) were strongly correlated with wet mount, gram's staining and culture findings.

Donders²⁹ conducted a study to assess vaginal flora on wet mount and gram stained specimens in all cases. They found that wet mount is quick to perform but Gram stain is performed more in routine. There was easier recognition of lactobacillary morph types on a wet mount than on gram stains which results in the loss of lactobacilli by the process of fixation or gram staining and recommended wet mount is cheaper and easier to perform for microscopy of vaginal smears rather than Gram staining. In this study it was also seen that wet mount was (41.7%) and gram stain (36%) positive in total cases.

In study by^{30,31} it was highlighted that Gram staining is gaining acceptance as diagnostic test of choice. It is simple for the physician who only has to smear a glass slide and allow it to air dry.

This study confirmed the common and established finding that gram method of staining is simple, inexpensive, sensitive, specific and reproducible way to diagnose³²

Our study reinforces the finding that vaginal cultures have the positive predictive value and is less than 60%. So cultures are not recommended^{33,19}. Vaginal cultures for *G. vaginalis* is not often the primary

laboratory test. The usefulness of these cultures is doubtful. In our study *G.vaginalis* could be recovered from 31.3% of women. So the incidental finding of *G.vaginalis* from a routine cultures should not be used unless clinical signs and/ or Gram staining shows its presence³³

In this study the presence of other organisms was not noted, as this study was oriented towards detection of *G.vaginalis*, and should not be interpreted as a study of the complete normal flora. This study also agrees with previous reports of^{34,35} that *G.vaginalis* can be found in vaginal secretions from some asymptomatic women. This study agrees with³⁵ that isolation and identification in routine laboratory is both time consuming and difficult. So diagnosis must usually be made on the basis of Amsel's criteria and on the characteristic microscopic appearance of wet mount and Gram stained smears of the discharge^{35,10}

CONCLUSION

The importance of an accurate, reproducible, and inexpensive laboratory method to diagnose bacterial vaginosis has increased with the recent association. Use of Amsel's clinical criteria and Gram staining especially in primary care unit and laboratories is recommended.

REFERENCES

- Gardner HL, Dukes CD. *Haemophilus Vaginalis*. Am J Obstet Gynecol 1955;69:962-76.
- Greenwood JR, Pickett MJ. Transfer of *Haemophilus vaginalis* Gardner and Dukes to a new genus. *Gardnerella*: *G. vaginalis* (Gardner and Dukes) comb. Nov Int J Syst Bacteriol 1980; 3:170-178.
- Donders GGG, Bulck BV, Caudron J, Londers L, Vereecken A, Spitz B. Relationship of bacterial vaginosis and mycoplasmas to the risk of spontaneous abortion. Am J Obstet Gynecol 2000;183:431-437.
- Egan ME, Lipsky MS. Diagnosis of vaginitis. Am Family Physician 2000;62:1095-1104.
- Adinkra P, Lamont RF. Adverse obstetric sequelae of bacterial vaginosis. Hospital Medicine 2000;61(7):475-477.
- Newton ER, Piper JM, Shain RN, Perdue ST, Peairs W. Predictors of the vaginal microflora. Am J Obstet Gynecol 2001;184:845-855.
- Schwebke JR. Asymptomatic bacterial vaginosis: Response to therapy. Am J Obstet Gynecol 2000;183:1434-1439.
- Koumans EH, Kendrick JS. Preventing adverse sequelae of bacterial vaginosis. A public health program and research agenda. Sex Tran Dis 2001;28(5):292-296.
- Morris M, Nicoll A, Simms I, Wilson J, Catchpole M. Bacterial vaginosis: a public health review. Br J Obstet Gynaecol 2001;108:439-450.
- Sweet RL. Gynecologic conditions and bacterial vaginosis: implications for the non-pregnant patient. Infect Dis Obstet Gynecol 2000;8:184-190.
- Helberg D, Nilsson S, Mardh PA. The diagnosis of bacterial vaginosis and vaginal flora changes. Arch Gynecol Obstet 2001;265:11-15.
- Sobel JD. Bacterial vaginosis. Annu Rev Med 2000;51:349-356.
- Morris M, Nicoll A, Simms I, Wilson J, Catchpole M. Bacterial vaginosis: a public health review. BJOG 2001;108:439-450.
- Schwebke JR. Gynecologic consequences of bacterial vaginosis. Obstet Gynecol Clin N Am 2003;30:685-694.
- Anderson MR, Klink K, Cohrssen A. Evaluation of vaginal complaints. JAMA 2004;291:1368-1379.
- Landers DV, Wiesenfeld HC, Heine RP, Krohn MA, Hillier SL. Predictive value of the clinical diagnosis of lower genital tract infection in women. Am J Obstet Gynecol 2004;190:1004-1010.
- Schaaf VM, Perez-Stable EJ, Borchardt K. The limited value of symptoms and signs in the diagnosis of vaginal infections. Arch Intern Med 1990;150:1929-1933.
- Amsel R, Totten PA, Spiegel CA, et al. Nonspecific vaginitis: Diagnostic criteria and microbial and epidemiologic associations. Am J Med 1983;74:14-22.
- Eschenbach DA, Hillier SL, Critchlow C, Stevens C, DeRouen T, Holmes KK. Diagnosis and clinical manifestations of bacterial vaginosis. Am J Obstet Gynecol 1988;158:819-828.
- Dadhwal V, Hariprasad R, Mittal S, Kapil A. Prevalence of bacterial vaginosis in pregnant women and predictive value of clinical diagnosis. Arch Gynecol Obstet 2009, [Epub ahead of print]
- Honest H, Bachmann LM, Knox EM, Gupta JK, Kleijnen J, Khan KS. The accuracy of various tests for bacterial vaginosis in predicting preterm birth: a systematic review. BJOG. 2004;111(5):409-22.
- Cohrssen A, Anderson M, Merrill A, McKee D. Reliability of the whiff test in clinical practice. J Am Board FamPract 2005;18:561-2.
- Cheesbrough M. District laboratory practice in tropical countries part 2, Cambridge University

- Press, Edinburgh, Cambridge, CB22, United Kingdom 2000;p.90-97.
24. Silonie S. Clue cell. *Indian J Dermatol Venerol Leprol* 2006; 72: 392 – 3.
25. Collee JG, Miles RS. Tests for identification of bacteria. In: Collee JG, Fraser AG, Duguid JP, Marmion BP, editors. *Mackie and McCartney Practical Medical Microbiology*. 13th ed. Edinburgh: Churchill Livingstone; 1989. p141-160.
26. Betty AF, Daniel FS, Alice SW. Laboratory considerations. In: Betty AF, Daniel FS, Alice SW, editors. *Bailey and Scott's Diagnostic Microbiology*. 12th ed. St. Louis, Missouri: Morby Elsevier; 2007. p.463-77.
27. 3M National Vaginitis Association News and Information. Archived Press Releases [online] accessed 21.07.2001. <http://www.vaginalinfections.com/archive.html>. pp.1-7.
28. Bernstein PS. (2000) Screening for bacterial vaginosis in pregnancy: A meta analysis. *Obstet Gynecol* 2000;95 (4 Suppl):57.
29. Donders GGG, Vereecken A, Dekeersmaecker A, VanBulck BV, Spitz B. Wet mount microscopy reflects functional vaginal lactobacillary flora better than gram stain. *J Clin Pathol* 2000;53: 308-313.
30. Rotimi VO, Yakubu Z, Abudu OO, Banjo TO. Direct Gram's stain of vaginal discharge as a means of diagnosing bacterial vaginosis. *J Med Microbiol* 1991;35:103-106.
31. MacDermontt RJ. Bacterial vaginosis *Br J Obstet Gynecol* 1995;102:92-94.
32. Mastrobattista JM, Bishop KD and Newton ER. Wet smear compared with gram stain Diagnosis of bacterial vaginosis in asymptomatic pregnant women. *Obstet Gynecol* 2000;96:504-506.
33. Spiegel CA, Amsel R and Holmes KK. Diagnosis of bacterial vaginosis by direct Gram stain of vaginal fluid. *J Clin Microbiol* 1983;18:170-177.
34. McCormack WM, Hayes CH, Rosner B, Evrard JR, Crockett VA, Alpert S, Zinner SH. Vaginal colonization with corynebacterium vaginale (*Haemophilus vaginalis*). *J Infec Dis* 1977; 136:740-744.
35. Pheifer TA, Forsyth PS, Durfee MA, Pollock HM, Holmes KK. Nonspecific vaginosis. Role of *Haemophilus vaginalis* and treatment with metronidazole. *N Engl J Med* 1978;298(26): 1429-1434.

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